

Review**Natural products' potential to maintain/ameliorate oral health: A review**

^{1,2*}Qamar, Z., ³Zeeshan, T., ⁴Al Dossary, O. B., ⁴Alanazi, T. A., ⁴Aldhuwayhi, J. N.,
⁴Alqarni, A. H. and ⁴Alshalan, A. M.

¹Department of O&MFS and Diagnostic Sciences, Faculty of Dentistry, Riyadh Elm University, Riyadh, Saudi Arabia

²Department of Oral Biology, Faculty of Dentistry, Liaquat College of Medicine and Dentistry, Karachi, Pakistan

³Department of Oral Biology and Biomedical Sciences, Faculty of Dentistry, University of Malaya,
 Kuala Lumpur, Malaysia

⁴Faculty of Dentistry, Vision Colleges, Riyadh, Saudi Arabia

Article history

Received:

9 December 2020

Received in revised form:

9 August 2021

Accepted:

9 September 2021

Keywords

dental caries,
 oral health,
 plant products,
 phenolic compounds

Abstract

Many diseases such as human dental caries result in the destruction of tooth structure; dental caries is an infectious disease leading to the destruction of tooth structure due to the acids produced on the fermentation of dietary carbohydrates by acidogenic bacteria. Bacteria colonise non-shedding oral surfaces, and produce lactic, acetic, and formic acids. Preserving tooth structure using fluoride in dental products may have its adverse effects on human health. The use of fluoride-containing dental products without supervision in children when the teeth are developing can lead to fluorosis. Therefore, the main aim of the present review was to identify natural oral healthcare products having minimum or no adverse effects in maintaining the integrity of tooth enamel. The active compounds observed in the natural plant products till date are polyphenolic compounds which contain antibacterial effects, and a potential to shift demineralisation to remineralisation. But their mechanism of action remain unclear. Therefore, further studies are needed to determine the effectiveness of these natural products and enhance their effect.

© All Rights Reserved

Introduction

Oral diseases are the most common chronic diseases (Frencken *et al.*, 2012) affecting the population. These diseases include dental caries, periodontal diseases, dental erosion, and developmental defects of enamel. Dental caries, also known as tooth decay, is a pathological process caused by the microorganisms in dental plaque resulting in localised destruction of the tissues at the tooth (Banerjee and Watson, 2015). The information regarding dental caries has been reported since 5000 B.C. Investigations are being carried out in order to determine its prevention and treatment. Several *in vivo* and *in vitro* studies have been reported regarding antibiotics that have potential to inhibit dental caries, but their long-term use leads to the alteration in human microflora, thus developing bacterial tolerance, diarrhoea, vomiting, and teeth staining, which further limit their use (Goldin and Gorbach, 1984; Dickinson and Surawicz, 2014; Gopinath *et al.*, 2014; Vennila *et al.*, 2014). In the past few decades,

various studies have been reported on fluoride and its anticaries effect by shifting the demineralisation and remineralisation balance. However, using fluoride in dental products may have its adverse effects. The use of fluoride-containing dental products in children without parental supervision, when teeth are developing, can lead to dental fluorosis (Seow, 2015). It may also cause burning mouth syndrome, sore tongue, nausea, vomiting, diarrhoea, increased saliva, stomach pain or cramp, muscle weakness, and seizures if swallowed in high dose (Choi *et al.*, 2012; Dey and Giri, 2015; Waugh *et al.*, 2016).

An oral healthcare product having minimum or no adverse effects is needed to maintain the integrity of tooth. Therefore, there is increasing interest on natural products which may have therapeutic uses in dentistry (Grosso *et al.*, 2008; Newman, 2008). Presently, investigations are being carried out using different foods and beverages such as tea, coffee, grape, propolis, shiitake (*Lentinula edodes*) mushroom, or traditional herbs (Spratt *et al.*, 2012).

*Corresponding author.
 Email: zeeshan.qamar@gmail.com

Most researchers have reported that the active compounds observed in natural products are polyphenol compounds, a substance containing a minimum of one aromatic ring with one or more hydroxyl group (Yoo *et al.*, 2011). Some of these compounds which are extracted from different natural products were showed to have bactericidal or bacteriostatic effect on oral biofilms (Ferrazzano *et al.*, 2011); whereas some others were observed to regulate the demineralisation and remineralisation of dental hard tissue (Chu *et al.*, 2007; Cheng *et al.*, 2008). In the present review, we summarised previous studies on the natural products and their active compounds, particularly for the prevention of dental caries.

Galla chinensis

Galla chinensis (GC), hypothetically an interesting agent for the prevention of dental caries, is produced from the leaves of *Rhus chinensis* Mill infested by the Chinese sumac aphids (*Melaphis chinensis* Bell). GC extracts have been reported to effectively inhibit demineralisation and promote remineralisation of dental hard tissue (Chu *et al.*, 2007; Cheng *et al.*, 2008; Zou *et al.*, 2008; Guo *et al.*, 2012). It was reported that the effect of fluoride in shifting demineralisation to remineralisation is enhanced with the presence of GC extracts (Cheng *et al.*, 2008). Huang *et al.* (2010) reported a potent effect of nanohydroxyapatite (HAp) and GC on remineralisation. To date, however, the mechanism of action of GC is unclear, but it has been suggested that the active compound of GC might act as a carrier in supplying calcium ions (Ca^{2+}) from the remineralising solution to the body of the early carious lesion (Chu *et al.*, 2007).

A hypothesis “enamel organic matrix - *Galla chinensis* - Ca” was proposed by Zhang *et al.* (2009a) based on the evidence of atomic force microscopy study. Morphological changes and increased roughness were observed on the carious enamel surface after treatment with the extracts of GC. An abundant number of nano-size elliptical particles were observed to be distributed over the lesion surface (Zhang *et al.*, 2009b). Furthermore, in order to analyse the morphological, chemical, and crystal characteristics, scanning of the remineralised early carious lesion using other techniques such as scanning electron microscopy, energy dispersive spectroscopy, and X-ray micro-diffraction was used, respectively (Zhang *et al.*, 2009b). GC is a complex

compound. Chu *et al.* (2007) extracted GC-B1 and GC-B2 by spectroscopic technique, and they were characterised later as gallic acid and methyl gallate, respectively. In an *in vitro* study, Cheng *et al.* (2008) compared the remineralising effect of GC extract with gallic acid using transverse microradiography (TMR). The results demonstrated that both of these compounds had the potential to reduce integrated mineral loss and lesion depth in comparison to the negative control. Further investigations displayed the potential of GC extract in remineralising the body of lesion, whereas gallic acid showed mineral deposition on the surface layer (Cheng *et al.*, 2008).

In order to observe the potential of GC extracts and gallic acids, researchers carried out investigations to determine the mechanism of remineralisation (Cheng *et al.*, 2009; 2010; Cheng and ten Cate, 2010). Gallic acids were observed to act as calcium carrier, and deposit the mineral on surface of the carious lesion. GC extract contained various polyphenol compounds and had the potential in preventing the mineral deposition on the surface layer of lesion, therefore transmitting calcium ions to the body of the carious lesion. Tang *et al.* (2015) further reported that gallic acids, in addition to up-regulating remineralisation, also have the potential to modulate the morphology and structure of the crystallites.

GC extracts are acidic in pH, which can demineralise enamel. Therefore, Huang *et al.* (2012) in an *in vitro* study determined the effect of pH on stability and demineralisation inhibition properties of GC extract. It was observed that GC extracts were unstable under neutral and alkaline pH levels. Furthermore, they reported that variation in pH levels of GC extract did not have an influence on demineralisation inhibition of dental enamel (Huang *et al.*, 2012).

Moreover, GC extract has also been reported to have potential in inhibiting dental biofilm formation. Cheng *et al.* (2010) in an *in vitro* experiment determined the effect of GC extracts on different stages of inoculated salivary biofilm, and observed a strong potential of the extract to inhibit microbial growth and acid production.

Xie *et al.* (2008) further analysed the potential of GC extract against four early colonisers of oral biofilm (*Streptococcus sanguis*, *S. mutans*, *Actinomyces naeslundii*, and *Lactobacillus rhamnosus*). The results showed a strong potential of GC extract in inhibiting multiple species of bacterial colonisation and demineralisation of dental enamel.

Propolis

Propolis has gained a vast research interest due to its antimicrobial effect against various microorganisms. Propolis is a compound comprises of flavonoids, organic acids, phenols, enzymes, vitamins, and minerals (Velikova *et al.*, 2000; Usia *et al.*, 2002). Koo *et al.* (2002b) were the first to report about propolis inhibitory effect against microbial growth and glucosyltransferase activity. Furthermore, Sonmez *et al.* (2005) determined the cytotoxicity of gingival fibroblasts, and reported strong antibacterial and non-cytotoxic activity of propolis.

To determine the mechanism of propolis, Duarte *et al.* (2006b) investigated the effect of different propolis compounds (ethanolic extract and hexane extract) on *S. mutans* biofilms and caries in rats. It was suggested that the anticaries effect of propolis is attributed to the high content of fatty acids which down-regulate acid production and tolerance by cariogenic microorganisms. Furthermore, Kouidhi *et al.* (2010) in an *in vitro* experiment reported a strong anticaries and antibiofilm potentials of Tunisian propolis ethanolic extract.

Magnolia bark

The bark of magnolia plant has been used as medication since the last 2,000 years (Watanabe *et al.*, 1983). Various researchers have reported that magnolol and honokiol, the two compounds of magnolia bark extract (MBE), have the potential to inhibit bacterial growth involved in dental caries (Chang *et al.*, 1998; Ho *et al.*, 2001; Greenberg *et al.*, 2007). On the basis of its anticariogenic properties, a group of researchers added MBE to xylitol chewing gum (Campus *et al.*, 2011). On 30 days randomised controlled intervention trial, a positive effect was observed on the oral health which included the decrease in salivary *S. mutans*, bleeding on probing, and plaque acidogenicity (Campus *et al.*, 2011).

Tea

Tea is a beverage consumed widely around the world, particularly in the Asian countries. In the United States, diet tea is considered a major source of flavonols and flavan-3-ols (Song and Chun, 2008). However, tea contains polyphenols whose types and levels depend on the technique from which the tea is being processed (Astill *et al.*, 2001).

The effectiveness of green tea extract (GTE) is attributed to its polyphenolic group (catechins, epigallocatechin-3-gallate, epigallocatechin,

epicatechin-3-gallate, and epicatechin) (Taylor *et al.*, 2005). Similarly, oolong tea extract (OTE) is also rich in polyphenolic groups (catechins and oligomerised catechins) (Balentine *et al.*, 1997). In contrast to GTE and OTE, active constituents in black tea are theaflavins (oligomers) and thearubigins (polymers). Various researchers have reported on the biologically active effect of these phenolic groups and theaflavins, particularly in the deterrence of dental caries and oral cancer (Yang, 1997; Yang *et al.*, 1999; 2000; 2002; Hamilton-Miller, 2001).

In various *in vivo* and *in vitro* studies, researchers have reported a potent effect of GTE against dental caries attributed to its polyphenolic group. It has been found to inhibit the bacterial colonisation of the cariogenic bacteria (*S. mutans*). Araghizadeh *et al.* (2013) in an *in vitro* study reported that other than anticariogenic effect, GTE also has the potential to inhibit periodontopathic bacterial colonisation. Furthermore, GTE has also been reported to decrease the incidence of pathological conditions such as oral cancer, stroke, cardiovascular disease, and obesity (Taylor *et al.*, 2005; Chacko *et al.*, 2010).

Another group of researchers (Awadalla *et al.*, 2011) suggested that rinsing with sugar-free solution of green tea can deter the growth of *S. mutans*, which is the cariogenic bacterium in both the saliva and dental plaque. Lee *et al.* (2004) suggested that the use of green tea leaves and brewed black tea could be suitable for the slow release of its active constituents catechins and theaflavins, respectively in order to prevent dental caries. The participants were asked to chew 2 g of green tea leaves or brewed black tea for the duration of 2 - 5 min. Later after rinsing, on performing high-performance liquid chromatography technique of saliva after 1 h, high concentrations of green tea catechins (GTC) (C max = 131.0 - 2.2 μ M) and black tea theaflavins (BTT) (C max = 1.8 - 0.6 μ M) were observed.

The GTE *Camellia* extract MJ (Taiyo Kagaku), which is also used as a food additive, contains over 1,500 ppm fluoride with approximately 0.06 g catechin/L. Whereas, GTE as a whole has been reported to contain 0.5 - 1 g of catechin/L. An *in situ* experiment was conducted by Suyama *et al.* (2011) using *Camellia* extract MJ in chewing gum, considering it as a rich natural source of fluoride. They observed an increase in acid resistance and remineralisation of dental enamel (Suyama *et al.*, 2011).

Nakahara *et al.* (1993) reported that the antiglycosyltransferase activity of OTE is attributed to its polymeric polyphenolic group. Later, Ooshima *et al.* (1993) in an *in vivo* study on rats evaluated an anticariogenic effect of OTE. The rats were infected with specific cariogenic bacteria, either *S. sobrinus* 6715 or *S. mutans* MT814R. Later, OTE was administered in the diet and drinking water which led to significant reduction in cariogenic bacteria and plaque deposition. In an *in vivo* study, Ooshima *et al.* (1994) reported that OTE had a potent effect in reducing the dental plaque in humans.

Matsumoto *et al.* (1999) reported that other than inhibiting cariogenic bacterial growth, OTE also had the potential to inhibit microbial colonisation on the tooth surface attributed to the polyphenolic group, which led to the reduction in cell surface hydrophobicity of *S. mutans*.

Grapes

Researchers have reported that grape (*Vitis vinifera*) and its seed extracts have an anticariogenic potential (Sarni-Manchado *et al.*, 1999; Daglia *et al.*, 2007a; Thimothe *et al.*, 2007). Duarte *et al.* (2006a) suggested that proanthocyanidins, a type of polyphenols, can inhibit dental caries as it has potential to reduce the activity of the surface-absorbed glycosyltransferase and F-ATPase; furthermore, it can also reduce acid production by cariogenic bacteria. There are a few types of grapes which contain phenolic compounds, and have the potential to reduce glycosyltransferase-B (GtfB) and glycosyltransferase-C (GtfC) activities of the cariogenic bacteria (*S. mutans*) (Yano *et al.*, 2012).

Xie *et al.* (2008) later assessed and reported that the additional potential of grape other than demineralisation inhibition is to promote remineralisation of an artificial carious lesion developed on the root of the tooth. It is proposed by various researchers that the seed extract has a potential for remineralisation by promoting deposition of mineral content on the superficial layer of the lesion (Kosasi *et al.*, 1989; Xie *et al.*, 2008). Later, on comparing the effect of grapes seed extracts and fluoride (Fluorinol®), separately, and in combination (2,000 µg/mL of seed extract and 10.2 mg/mL fluoride), the antiplaque and antioxidant potency of grape seed extracts was observed to increase when used in combination with fluoride (Furiga *et al.*, 2014).

The phenolic content in the wines produced by grape extract have been suggested to inhibit bacterial growth. The phenolic compounds in wine, also in grape and pomace extracts, have been reported to have bactericidal effect on the cariogenic bacteria (*Streptococcus* spp.) (Thimothe *et al.*, 2007). Munoz-Gonzalez *et al.* (2014) determined the effect of red wine and alcohol-free red wine on biofilm model of the supra-gingival plaque, and reported that both of them possessed strong antimicrobial effect, particularly against *F. nucleatum* and *S. oralis*.

Coffee

Other than tea, other most commonly consumed drink globally is coffee which has been reported to be effective against various diseases including dental caries (Daglia *et al.*, 2007b; Antonio *et al.*, 2011; 2012). The effectiveness of coffee against bacteria is due to the presence of polyphenols (Koo *et al.*, 2002a; Yatsuda *et al.*, 2005; Farah *et al.*, 2006). Antonio *et al.* (2011; 2012) observed bactericidal effect of coffee against *S. mutans* in addition to demineralisation inhibition potential of biofilm-coated dental enamel. The bactericidal effect of coffee is attributed to the bacterial lysis followed by calcium release. Meckelburg *et al.* (2014) also reported that the consumption of coffee could be effective in inhibiting bacterial growth.

Cacao bean

The major compound of chocolate from cacao beans is polyphenols which have antiglycosyltransferase potential. Ooshima *et al.* (2000b) in an *in vitro* study reported anticariogenic potential, but not strong enough to reduce the cariogenic activity of sucrose. They further suggested that cocoa has the potential to become a novel anticariogenic prophylactic agent (Ooshima *et al.*, 2000a).

Hesperidin

Hesperidin is a natural product rich in citrus flavonoids, and has the potential to preserve collagen matrix of the bovine dentinal matrix against the proteolytic degradation. Liu *et al.* (2004) reported the mechanism of action and interaction of hesperidin with the collagenous and non-collagenous proteins, which resulted in stabilising the collagen matrix and promoting remineralisation of the human dental hard tissue.

Hiraishi *et al.* (2011) determined that hesperidin had the capability to reduce demineralisation from acids, and increase remineralisation process. Later, a group of scientists studied the effect of hesperidin on mineral loss and depth of lesion which displayed the potential of the product to inhibit demineralisation and promote remineralisation under fluoride-free conditions (Islam *et al.*, 2012).

Poly- γ -glutamic acid

Poly- γ -glutamic acid (PGGA) is a naturally occurring homopolyamide containing D- and L-forms of glutamic acid units, and connected via amide linkages (Qamar *et al.*, 2016). These linkages are moulded between the α -amino and γ -carboxyl group. Qamar *et al.* (2016) suggested that PGGA, being a highly viscous material, had the potential for coating the enamel surface of the tooth by forming a protective layer. The α -COOH group in PGGA molecule is free, and may promote binding of protective coating layer to the enamel surface, thus inhibiting the dissolution of the enamel HAP.

Furthermore, as a mechanism of action, the anionic α -COOH group has been proposed as capable to bind cationic entity of other molecule or biopolymer, or can behave or remain as a free carboxylic acid. Therefore, PGGA can dissolve particularly Ca and Mg compounds to form a stable ionic complexes (Qamar *et al.*, 2016; 2019).

Conclusion

It can be concluded from the present review that natural products do have potent effect in inhibiting dental caries, but not without some drawbacks. Firstly, they have a weaker effect than traditional antimicrobials or fluoride-containing chemotherapeutic agents. Secondly, the active compounds in various natural products are still unclear; therefore, further studies are required to determine the active compounds in natural products with cariogenic potential, and modify them for stronger potential.

References

- Antonio, A. G., Iorio, N. L., Farah, A., Netto dos Santos, K. R. and Maia, L. C. 2012. Effect of *Coffea canephora* aqueous extract on microbial counts in *ex vivo* oral biofilms: a case study. *Planta Medica* 78(8): 755-760.
- Antonio, A. G., Iorio, N. L., Pierro, V. S., Candreva, M. S., Farah, A., dos Santos, K. R. and Maia, L. C. 2011. Inhibitory properties of *Coffea canephora* extract against oral bacteria and its effect on demineralisation of deciduous teeth. *Archives of Oral Biology* 56(6): 556-564.
- Araghizadeh, A., Kohanteb, J. and Fani, M. M. 2013. Inhibitory activity of green tea (*Camellia sinensis*) extract on some clinically isolated cariogenic and periodontopathic bacteria. *Medical Principles and Practice* 22(4): 368-372.
- Astill, C., Birch, M. R., Dacombe, C., Humphrey, P. G. and Martin, P. T. 2001. Factors affecting the caffeine and polyphenol contents of black and green tea infusions. *Journal of Agricultural and Food Chemistry* 49(11): 5340-5347.
- Awadalla, H. I., Ragab, M. H., Bassuoni, M. W., Fayed, M. T. and Abbas, M. O. 2011. A pilot study of the role of green tea use on oral health. *International Journal of Dental Hygiene* 9(2): 110-116.
- Balentine, D. A., Wiseman, S. A. and Bouwens, L. C. M. 1997. The chemistry of tea flavonoids. *Critical Reviews in Food Science and Nutrition* 37(8): 693-704.
- Banerjee, A. and Watson, T. F. 2015. Dental hard tissue pathologies, aetiology, and their clinical manifestation. In Banerjee, A. and Watson, T. F. (eds). *Pickard's Guide to Minimally Invasive Operative Dentistry*, p. 1-15. United Kingdom: Oxford Scholarship Online.
- Campus, G., Cagetti, M. G., Cocco, F., Sale, S., Sacco, G., Strohmeier, L. and Lingström, P. 2011. Effect of a sugar-free chewing gum containing magnolia bark extract on different variables related to caries and gingivitis: a randomized controlled intervention trial. *Caries Research* 45(4): 393-399.
- Chacko, S. M., Thambi, P. T., Kuttan, R. and Nishigaki, I. 2010. Beneficial effects of green tea: a literature review. *Chinese Medical Journal* 5: article no. 13.
- Chang, B., Lee, Y., Ku, Y., Bae, K. and Chung, C. 1998. Antimicrobial activity of magnolol and honokiol against periodontopathic microorganisms. *Planta Medica* 64(4): 367-369.

- Cheng, L. and ten Cate, J. M. 2010. Effect of *Galla chinensis* on the *in vitro* remineralization of advanced enamel lesions. *International Journal of Oral Science* 2(1): 15-20.
- Cheng, L., Li, J., Hao, Y. and Zhou, X. 2008. Effect of compounds of *Galla chinensis* and their combined effects with fluoride on remineralization of initial enamel lesion *in vitro*. *Journal of Dentistry* 36(5): 369-373.
- Cheng, L., Li, J., Hao, Y. and Zhou, X. 2010. Effect of compounds of *Galla chinensis* on remineralization of enamel surface *in vitro*. *Archives of Oral Biology* 55(6): 435-440.
- Cheng, L., Li, J. Y., Huang, S. and Zhou, X. D. 2009. Effect of *Galla chinensis* on enhancing remineralization of enamel crystals. *Biomedical Materials* 4(3): article ID 034103.
- Choi, A. L., Sun, G. F., Zhang, Y. and Grandjean, P. 2012. Developmental fluoride neurotoxicity: a systematic review and meta-analysis. *Environmental Health Perspectives* 120(10): 1362-1368.
- Chu, J. P., Li, J. Y., Hao, Y. Q. and Zhou, X. D. 2007. Effect of compounds of *Galla chinensis* on remineralisation of initial enamel carious lesions *in vitro*. *Journal of Dentistry* 35(5): 383-387.
- Daglia, M., Papetti, A., Grisoli, P., Aceti, C., Dacarro, C. and Gazzani, G. 2007a. Antibacterial activity of red and white wine against oral streptococci. *Journal of Agricultural and Food Chemistry* 55(13): 5038-5042.
- Daglia, M., Papetti, A., Grisoli, P., Aceti, C., Spini, V., Dacarro, C. and Gazzani, G. 2007b. Isolation, identification, and quantification of roasted coffee antibacterial compounds. *Journal of Agricultural and Food Chemistry* 55(25): 10208-10213.
- Dey, S. and Giri, B. 2015. Fluoride fact on human health and health problems: a review. *Medical and Clinical Reviews* 2(1): article no. 2.
- Dickinson, B. and Surawicz, C. M. 2014. Infectious diarrhea: an overview. *Current Gastroenterology Reports* 16(8): article no. 399.
- Duarte, S., Rosalen, P. L., Hayacibara, M. F., Cury, J. A., Bowen, W. H., Marquis, R. E., ... and Koo, H. 2006b. The influence of a novel propolis on mutans streptococci biofilms and caries development in rats. *Archives of Oral Biology* 51(1): 15-22.
- Duarte, S., Gregoire, S., Singh, A. P., Vorsa, N., Schaich, K., Bowen, W. H. and Koo, H. 2006a. Inhibitory effects of cranberry polyphenols on formation and acidogenicity of *Streptococcus mutans* biofilms. *FEMS Microbiology Letters* 257(1): 50-56.
- Farah, A., de Paulis, T., Moreira, D. P., Trugo, L. C. and Martin, P. R. 2006. Chlorogenic acids and lactones in regular and water-decaffeinated arabica coffees. *Journal of Agricultural and Food Chemistry* 54(2): 374-381.
- Ferrazzano, G. F., Amato, I., Ingenito, A., Zarrelli, A., Pinto, G. and Pollio, A. 2011. Plant polyphenols and their anti-cariogenic properties: a review. *Molecules* 16(2): 1486-1507.
- Frencken, J. E., Peters, M. C., Manton, D. J., Leal, S. C., Gordan, V. V. and Eden, E. 2012. Minimal intervention dentistry for managing dental caries - a review. *International Dental Journal* 62(5): 223-243.
- Furiga, A., Roques, C. and Badet, C. 2014. Preventive effects of an original combination of grape seed polyphenols with amine fluoride on dental biofilm formation and oxidative damage by oral bacteria. *Journal of Applied Microbiology* 116(4): 761-771.
- Goldin, B. R. and Gorbach, S. L. 1984. Alterations of the intestinal microflora by diet, oral antibiotics, and *Lactobacillus*: decreased production of free amines from aromatic nitro compounds, azo dyes, and glucuronides. *Journal of the National Cancer Institute* 73(3): 689-695.
- Gopinath, S., Lichtman, J. S., Bouley, D. M., Elias, J. E. and Monack, D. M. 2014. Role of disease-associated tolerance in infectious superspreaders. *Proceedings of the National Academy of Sciences of the United States of America* 111(44): 15780-15785.
- Greenberg, M., Urnezis, P. and Tian, M. 2007. Compressed mints and chewing gum containing magnolia bark extract are effective against bacteria responsible for oral malodor. *Journal of Agricultural and Food Chemistry* 55(23): 9465-9469.
- Grosso, F. C., Bergamaschi, C., Cogo, K., Franz-Montan, M., Motta, R. H. and de Andrade, E. D. 2008. Use of phytotherapy in dentistry. *Phytotherapy Research* 22(8): 993-998.

- Guo, B., Que, K. H., Yang, J., Wang, B., Liang, Q. Q. and Xie, H. H. 2012. Effect of *Galla chinensis* on the remineralization of two bovine root lesions morphous *in vitro*. International Journal of Oral Sciences 4(3): 152-156.
- Hamilton-Miller, J. M. 2001. Anti-cariogenic properties of tea (*Camellia sinensis*). Journal of Medical Microbiology 50(4): 299-302.
- Hiraishi, N., Sono, R., Islam, M. S., Otsuki, M., Tagami, J. and Takatsuka, T. 2011. Effect of hesperidin *in vitro* on root dentine collagen and demineralization. Journal of Dentistry 39(5): 391-396.
- Ho, K. Y., Tsai, C. C., Chen, C. P., Huang, J. S. and Lin, C. C. 2001. Antimicrobial activity of honokiol and magnolol isolated from *Magnolia officinalis*. Phytotherapy Research 15(2): 139-141.
- Huang, S., Gao, S., Cheng, L. and Yu, H. 2010. Combined effects of nano-hydroxyapatite and *Galla chinensis* on remineralisation of initial enamel lesion *in vitro*. Journal of Dentistry 38(10): 811-819.
- Huang, X., Cheng, L., Exterkate, R. A. M., Liu, M., Zhou, X., Li, J. and Ten Cate, J. M. 2012. Effect of pH on *Galla chinensis* extract's stability and anti-caries properties *in vitro*. Archives of Oral Biology 57(8): 1093-1099.
- Islam, S. M., Hiraishi, N., Nassar, M., Sono, R., Otsuki, M., Takatsuka, T., ... and Tagami, J. 2012. *In vitro* effect of hesperidin on root dentin collagen and de/re-mineralization. Dental Materials Journal 31(3): 362-367.
- Koo, H., Cury, J. A., Rosalen, P. L., Ambrosano, G. M., Ikegaki, M. and Park, Y. K. 2002a. Effect of a mouthrinse containing selected propolis on 3-day dental plaque accumulation and polysaccharide formation. Caries Research 36(6): 445-448.
- Koo, H., Rosalen, P. L., Cury, J. A., Park, Y. K. and Bowen, W. H. 2002b. Effects of compounds found in propolis on *Streptococcus mutans* growth and on glucosyltransferase activity. Antimicrobial Agents and Chemotherapy 46(5): 1302-1309.
- Kosasi, S., Hart, L. A., van Dijk, H. and Labadie, R. P. 1989. Inhibitory activity of *Jatropha multifida* latex on classical complement pathway activity in human serum mediated by a calcium-binding proanthocyanidin. Journal of Ethnopharmacology 27(1-2): 81-89.
- Kouidhi, B., Zmantar, T. and Bakhrouf, A. 2010. Anti-cariogenic and anti-biofilms activity of Tunisian propolis extract and its potential protective effect against cancer cells proliferation. Anaerobe 16(6): 566-571.
- Lee, M. J., Lambert, J. D., Prabhu, S., Meng, X., Lu, H., Maliakal, P., ... and Yang, C. S. 2004. Delivery of tea polyphenols to the oral cavity by green tea leaves and black tea extract. Cancer Epidemiology, Biomarkers and Prevention 13(1): 132-137.
- Liu, Y., Li, Y., Liu, S., Li, J. and Yao, S. 2004. Monitoring the self-assembly of chitosan/glutaraldehyde/cysteamine/Au-colloid and the binding of human serum albumin with hesperidin. Biomaterials 25(26): 5725-5733.
- Matsumoto, M., Minami, T., Sasaki, H., Sobue, S., Hamada, S. and Ooshima, T. 1999. Inhibitory effects of oolong tea extract on caries-inducing properties of mutans streptococci. Caries Research 33(6): 441-445.
- Meckelburg, N., Pinto, K. C., Farah, A., Iorio, N. L., Pierro, V. S., dos Santos, K. R., ... and Antonio, A. G. 2014. Antibacterial effect of coffee: calcium concentration in a culture containing teeth/biofilm exposed to *Coffea canephora* aqueous extract. Letters in Applied Microbiology 59(3): 342-347.
- Munoz-Gonzalez, I., Thurnheer, T., Bartolomé, B. and Moreno-Arribas, M. V. 2014. Red wine and oenological extracts display antimicrobial effects in an oral bacteria biofilm model. Journal of Agricultural and Food Chemistry 62(20): 4731-4737.
- Nakahara, K., Kawabata, S., Ono, H., Ogura, K., Tanaka, T., Ooshima, T. and Hamada, S. 1993. Inhibitory effect of oolong tea polyphenols on glycosyltransferases of mutans streptococci. Applied and Environmental Microbiology 59(4): 968-973.
- Newman, D. J. 2008. Natural products as leads to potential drugs: an old process or the new hope for drug discovery? Journal of Medicinal Chemistry 51(9): 2589-2599.
- Ooshima, T., Minami, T., Aono, W., Izumitani, A., Sobue, S., Fujiwara, T., ... and Hamada, S. 1993. Oolong tea polyphenols inhibit experimental dental caries in SPF rats infected with mutans streptococci. Caries Research 27(2): 124-129.

- Ooshima, T., Minami, T., Aono, W., Tamura, Y. and Hamada, S. 1994. Reduction of dental plaque deposition in humans by oolong tea extract. *Caries Research* 28(3): 146-149.
- Ooshima, T., Osaka, Y., Sasaki, H., Osawa, K., Yasuda, H. and Matsumoto, M. 2000a. Cariostatic activity of cacao mass extract. *Archives of Oral Biology* 45(9): 805-808.
- Ooshima, T., Osaka, Y., Sasaki, H., Osawa, K., Yasuda, H., Matsumura, M., ... and Matsumoto, M. 2000b. Caries inhibitory activity of cacao bean husk extract in *in vitro* and animal experiments. *Archives of Oral Biology* 45(8): 639-645.
- Qamar, Z., Haji Abdul Rahim, Z. B., Neon, G. S., Chew, H. P. and Zeeshan, T. 2019. Effectiveness of poly- γ -glutamic acid in maintaining enamel integrity. *Archives of Oral Biology* 106: article ID 104482.
- Qamar, Z., Haji Abdul Rahim, Z. B., Chew, H. P. and Fatima, T. 2016. Poly- γ -glutamic acid a substitute of salivary protein statherin? *Journal of the Chemical Society of Pakistan* 38(4): 730-736.
- Sarni-Manchado, P., Cheynier, V. and Moutounet, M. 1999. Interactions of grape seed tannins with salivary proteins. *Journal of Agricultural and Food Chemistry* 47(1): 42-47.
- Seow, W. K. 2015. Dental enamel defects in the primary dentition: prevalence and etiology. In Drummond, K. B. and Kilpatrick, N. (eds). *Planning and Care for Children and Adolescents with Dental Enamel Defects: Etiology, Research and Contemporary Management*, p. 1-14. Berlin: Springer.
- Song, W. O. and Chun, O. K. 2008. Tea is the major source of flavan-3-ol and flavonol in the US diet. *The Journal of Nutrition* 138(8): 1543S-1547S.
- Sonmez, S., Kirilmaz, L., Yucesoy, M., Yücel, B. and Yilmaz, B. 2005. The effect of bee propolis on oral pathogens and human gingival fibroblasts. *Journal of Ethnopharmacology* 102(3): 371-376.
- Spratt, D. A., Daglia, M., Papetti, A., Stauder, M., O'Donnell, D., Ciric, L., ... and Wilson, M. 2012. Evaluation of plant and fungal extracts for their potential antigingivitis and anticaries activity. *BioMed Research International* 2012: article ID 510198.
- Suyama, E., Tamura, T., Ozawa, T., Suzuki, A., Iijima, Y. and Saito, T. 2011. Remineralization and acid resistance of enamel lesions after chewing gum containing fluoride extracted from green tea. *Australian Dental Journal* 56(4): 394-400.
- Tang, B., Yuan, H., Cheng, L., Zhou, X., Huang, X. and Li, J. 2015. Effects of gallic acid on the morphology and growth of hydroxyapatite crystals. *Archives of Oral Biology* 60(1): 167-173.
- Taylor, P. W., Hamilton-Miller, J. M. and Stapleton, P. D. 2005. Antimicrobial properties of green tea catechins. *Food Science and Technology Bulletin* 2: 71-81.
- Thimothe, J., Bonsi, I. A., Padilla-Zakour, O. I. and Koo, H. 2007. Chemical characterization of red wine grape (*Vitis vinifera* and *Vitis* interspecific hybrids) and pomace phenolic extracts and their biological activity against *Streptococcus mutans*. *Journal of Agricultural and Food Chemistry* 55(25): 10200-10207.
- Usia, T., Banskota, A. H., Tezuka, Y., Midorikawa, K., Matsushige, K. and Kadota, S. 2002. Constituents of Chinese propolis and their antiproliferative activities. *Journal of Natural Products* 65(5): 673-676.
- Velikova, M., Bankova, V., Marcucci, M. C., Tsvetkova, I. and Kujumgiev, A. 2000. Chemical composition and biological activity of propolis from Brazilian Meliponinae. *Zeitschrift für Naturforschung C* 55(9-10): 785-789.
- Vennila, V., Madhu, V., Rajesh, R., Ealla, K. K., Velidandla, S. R. and Santoshi, S. 2014. Tetracycline-induced discoloration of deciduous teeth: case series. *Journal of International Oral Health* 6(3): 115-119.
- Watanabe, K., Watanabe, H., Goto, Y., Yamaguchi, M., Yamamoto, N. and Hagino, K. 1983. Pharmacological properties of magnolol and honokiol extracted from *Magnolia officinalis*: central depressant effects. *Planta Medica* 49(2): 103-108.
- Waugh, D. T., Potter, W., Limeback, H. and Godfrey, M. 2016. Risk assessment of fluoride intake from tea in The Republic of Ireland and its implications for public health and water fluoridation. *International Journal of Environmental Research and Public Health* 13(3): article no. 259.

- Xie, Q., Bedran-Russo, A. K. and Wu, C. D. 2008. *In vitro* remineralization effects of grape seed extract on artificial root caries. *Journal of Dentistry* 36(11): 900-906.
- Yang, C. S. 1997. Inhibition of carcinogenesis by tea. *Nature* 389: 134-135.
- Yang, C. S., Chung, J. Y., Yang, G. Y., Li, C., Meng, X. and Lee, M. J. 2000. Mechanisms of inhibition of carcinogenesis by tea. *Biofactors* 13(1-4): 73-79.
- Yang, C. S., Maliakal, P. and Meng, X. 2002. Inhibition of carcinogenesis by tea. *Annual Review of Pharmacology and Toxicology* 42: 25-54.
- Yang, C. S., Kim, S., Yang, G. Y., Lee, M. J., Liao, J., Chung, J. Y. and Ho, C. T. 1999. Inhibition of carcinogenesis by tea: bioavailability of tea polyphenols and mechanisms of actions. *Proceedings of the Society for Experimental Biology and Medicine* 220(4): 213-217.
- Yano, A., Kikuchi, S., Takahashi, T., Kohama, K. and Yoshida, Y. 2012. Inhibitory effects of the phenolic fraction from the pomace of *Vitis coignetiae* on biofilm formation by *Streptococcus mutans*. *Archives of Oral Biology* 57(6): 711-719.
- Yatsuda, R., Rosalen, P. L., Cury, J. A., Murata, R. M., Rehder, V. L., Melo, L. V. and Koo, H. 2005. Effects of *Mikania* genus plants on growth and cell adherence of mutans streptococci. *Journal of Ethnopharmacology* 97(2): 183-189.
- Yoo, S., Murata, R. M. and Duarte, S. 2011. Antimicrobial traits of tea- and cranberry-derived polyphenols against *Streptococcus mutans*. *Caries Research* 45(4): 327-335.
- Zhang, L. L., Li, J. Y., Zhou, X. D., Cui, F. Z. and Li, W. 2009a. Effects of *Galla chinensis* on the surface topography of initial enamel carious lesion: an atomic force microscopy study. *Scanning* 31(5): 195-203.
- Zhang, L. L., Li, J. Y., Zhou, X. D., Cui, F. Z. and Li, W. 2009b. Chemical and crystallographic study of remineralized surface on initial carious enamel treated with *Galla chinensis*. *Scanning* 31(6): 236-245.
- Zou, L., Zhang, L., Li, J., Hao, Y., Cheng, L., Li, W. and Zhou, X. 2008. Effect of *Galla chinensis* extract and chemical fractions on demineralization of bovine enamel *in vitro*. *Journal of Dentistry* 36(12): 999-1004.